Inhibition of Sodium- and Potassium-Dependent Adenosine Triphosphatase by Cardenolide Alkylating Agents¹

HANLEY N. ABRAMSON* AND JUNG-SOOK K. CHO

College of Pharmacy, Wayne State University, Detroit, Michigan 48202

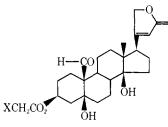
Received August 10, 1970

Four bromoacetylated cardenolides, namely, digitoxigenin 3-bromoacetate (2a), 15α -bromoacetoxydigitoxigenin 3-acetate (2b), digoxigenin 3,12-dibromoacetate (2c), and Δ^{14} -anhydrodigitoxigenin 3-bromoacetate (3a), were prepared and tested for their ability to inhibit Na- and K-ATPase from guinea pig brain. Three of these compounds, 2a, 2b, and 2c, inhibited the enzyme to approximately the same extent. Compound 3a had significant irreversible activity in spite of the fact that the corresponding 3-acetate (3b) did not appreciably inhibit the enzyme in a reversible fashion.

The naturally occurring cardiotonic steroids have long been accepted in medicine for their potent effects on the human heart. Investigations² into the mode of action of these compounds have demonstrated that they act as inhibitors of active ion transport. More specifically, it has been shown³ that this interference with ion transport is a result of inhibition of Na⁺- and K⁺-dependent ATPase (also called transport ATPase) which, by catalyzing the cleavage of ATP to ADP, provides the energy required for active ion transport.

The dephosphorylation of a postulated⁴ acyl phosphate ester is K⁺ dependent and this stage is inhibited by the cardiotonic steroid, ouabain.⁵ This inhibition is believed to be a result of displacement of K^+ from its attachment to the enzyme by the steroid in a competitive-type reaction.^{3,6} Portius and Repke⁷ have suggested that the CO group of the unsaturated lactone, characteristic of the cardiac steroids, forms an H bond with an OH of the phosphoric acid residue in the postulated phosphorylated enzyme intermediate. This H bonding prevents access of K^+ to the phosphorylated enzyme and also permits free rotation of the steroid molecule so that its correct face comes into close relationship with the complementary enzyme surface. Alternatively, Repke⁸ has proposed that the H bond involves a free OH, NH₂, or SH group in the enzyme. Likewise, Glynn⁹ has postulated that the unsaturated lactone ring may interact with SH or other nucleophilic groups at the K⁺ transport site, although Hoffman¹⁰ has adduced evidence that these steroids may actually inhibit in an allosteric manner rather than by direct competition with K^+ .

In an effort to further elucidate the molecular mechanism of action of the cardiotonic steroids Kupchan, *et al.*,¹¹ used the concept of active-site-directed inhibition.¹² They found that strophanthidin 3-iodoacetate and bromoacetate (1, X = I and Br, respectively) are both active-site-directed inhibitors of Na- and K-



ATPase obtained from guinea pig brain and also exhibit high cardiotonic activity in the guinea pig atrial perparation.

As part of a study to further define the nature of the inter-action between the cardiotonic steroids and Naand K-ATPase we have prepared 4 bromoacetylated cardenolide derivatives—*viz.*, digitoxigenin 3-bromoacetate (**2a**), 15α -bromoacetoxydigitoxigenin 3-acetate (**2b**), digoxigenin 3,12-dibromoacetate (**2c**), and Δ^{14} anhydrodigitoxigenin 3-bromoacetate (**3a**)—in order to test the ability of each to irreversibly inhibit Naand K-ATPase.

Synthesis.—Digitoxigenin acetate (2d) served as the starting material in the synthesis of 15α -bromoacetoxydigitoxigenin 3-acetate (2b). Dehydration of 2d to give Δ^{14} -anhydrodigitoxigenin 3-acetate (3b) was accomplished using the procedure of Bach, *et al.*¹³ Treatment of 3b with *m*-chloroperbenzoic acid gave the $14,15\alpha$ -epoxide 4 which was hydrolyzed with HClO₄¹⁴ to yield 15α -hydroxydigitoxigenin 3-acetate (2e). Bromoacetylation of 2e to give 2b was achieved by heating with BrCH₂COBr in dioxane-pyridine at 70° for 16 hr.

The placement of the bromoacetyl group at position 3 of digitoxigenin (5) was carried out at room temp using $BrCH_2COBr$ in dioxane-pyridine. It was found

⁽¹⁾ This work was supported by a grant from the Michigan Heart Association. Taken from the dissertation submitted by J.-S. K. Cho, in partial fulfillment of the requirements for a Master of Science degree. Wayne State University. Detroit. Mich., 1970.

^{(2) (}a) K. S. Lee, D. M. Yu. D. I. Lee, and R. Burstein, J. Pharmacol. Exp. Ther., 132, 139 (1961);
(b) W. Klaus, G. Kuschinsky, and H. Lullman. Naunyn-Schmiedebergs Arch. Exp. Path. Pharmakol., 242, 480 (1962);
(c) I. M. Glynn, Pharmacol. Rev., 16, 381 (1964).

^{(3) (}a) P. S. Caldwell and R. O. Keynes, J. Physiol., 148, 8 (1959); (b)
J. D. Judah and K. Ahmed, J. Cell. Comp. Physiol., 64, 355 (1964); (c)
D. E. Duggan and R. M. Noll. Arch. Biochem., 109, 388 (1965).

^{(4) (}a) L. E. Hokin and A. Yoda. Proc. Nat. Acad. Sci. U. S., 52, 454 (1964);
(b) L. E. Hokin, P. S. Sastry, P. R. Galsworthy, and A. Yoda. *ibid.*, 54, 177 (1965);
(c) K. Nagano, T. Kanazawa, N. Mizuno, Y. Tashima, T. Nakao, and M. Nakao. Biochem. Biophys. Res. Commun., 19, 759 (1965);
(d) D. A. Hems and R. Rodnight, Biochem. J., 101, 516 (1966);
(e) H. Bader, A. K. Sen, and R. L. Post, Biochim. Biophys. Acta. 118, 106 (1966);
(f) A. Kahlenberg, P. R. Galsworthy, and L. E. Hokin. Arch. Biochem. Biophys., 126, 331 (1968);

⁽⁵⁾ J. S. Charnock and R. L. Post, Nature (London), 208, 471 (1965).
(6) (a) L. J. Opit and J. S. Charnock, *ibid.*, 199, 910 (1963): (b) J. C.

⁽b) (a) L. b. Opti and b. S. Charnock, *ibid.*, **199**, 910 (1963): (b) Skou, *Physiol. Rev.*, **45**, 596 (1965).

⁽⁷⁾ H. J. Portius and K. Repke. Arzneim.-Forsch., 14, 1073 (1964).

⁽⁸⁾ K. Repke. Proc. Int. Pharmacol. Meet. 2nd. 1963, 77 (1965).

⁽⁹⁾ I. M. Glynn. J. Physiol.. 136, 148 (1957).

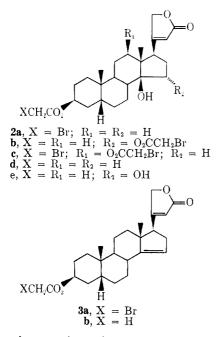
⁽¹⁰⁾ J. F. Hoffman, Amer. J. Med.. 41, 666 (1966).

 ^{(11) (}a) L. E. Hokin, M. Mokotoff, and S. M. Kupchan. Proc. Nat. Acad.
 Sci. U. S., 55, 797 (1966): (b) S. M. Kupchan, M. Mokotoff, R. S. Sandhu,
 and L. E. Hokin, J. Med. Chem., 10, 1025 (1967).

⁽¹²⁾ B. R. Baker. "Design of Active-Site-Directed Irreversible Enzyme Inhibitors. The Organic Chemistry of the Enzymic Active Site," Wiley, New York, N. Y., 1967.

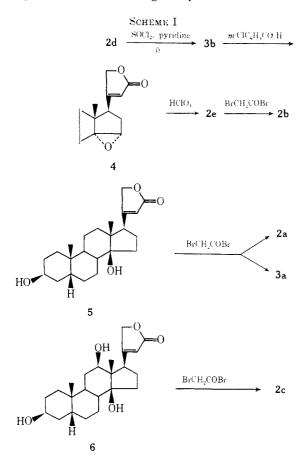
⁽¹³⁾ G. Bach, J. Capitaine, and C. H. R. Engel. Can. J. Chem., 46, 7333 (1968).

⁽¹⁴⁾ M. Okada and M. Hasunuma. Yakugaku Zasshi. 85, 822 (1965).



that when the reaction mixture was allowed to stand for 1.5 hr, 2a was the major product, but when the reaction time was extended to 3 hr, 3a was the predominant component of the reaction mixture.

When digoxigenin (6) was heated in dioxane-pyridine with BrCH₂COBr at 75°, 2c was obtained in 56% yield after purification on a silica gel tlc plate.



Biochemical Studies.—The inhibition studies were carried out using a Na- and K-ATPase preparation obtained from guinea pig brain. The procedure of Kupchan, *et al.*,^{11b} for obtaining the preparation was followed. The quantity of protein in the preparation was estimated by the method of Lowry, *et al.*,¹⁵ and the activity of the ATPase in both reversible and irreversible inhibition studies was determined by the quantity of inorganic phosphate liberated from the ATP substrate following the procedure used by Kupchan, *et al.*,^{11b} for the evaluation of strophanthidin 3-haloacetates.

Table I shows the effect of the cardenolide derivatives

TABLE I Na- and K-ATPASE INHIBITION STUDIES

	-% of control activity		-% of enzyme inhibition-,	
No. ^a	Un• washed	Washed	Reversible	Irre. versible
2a	7.1	18.9	11.8	81.1
2b	13.3	19.6	6.3	80.4
2c	5.8	15.5	9.7	84.5
2d	67.5	100	32.5	0.0
2e	98.0	104	6.0	-4.0
3a	20.0	29.7	9.7	70.3
3b	96.6	96.7	0.1	3.3
4	93.7	108	14.3	-8.0
5	53.5	105	51.5	-5.0
6	32.3	85.8	53.5	14.2

 $^{\rm a}$ All incubations were carried out with a final cardenolide conen of $10^{-4}\,M_{\odot}$

on brain Na- and K-ATPase. Na- and K-ATPase activity in the presence of cardenolide is expressed as a per cent of the activity of the control sample incubated without cardenolide. Irreversible inhibition of the Na- and K-ATPase was demonstrated by failure to regain the control activity after washing. The washing procedure brought the enzyme activity back to that of the control when the inhibition was of a reversible nature. The gain in activity after washing is the per cent of reversible inhibition. The activity not regained after washing is the per cent of irreversible inhibition.

Discussion

Digitoxigenin (5) and digoxigenin (6) inhibited the ATPase only before washing and this inhibition was reversible. This agrees well with the results of Repke and Portius¹⁶ who showed that digitoxigenin inhibited Na- and K-ATPase by 50% at $1.2 \times 10^{-4} M$. Digitoxigenin 3-acetate (2d) also showed reversible inhibition in our study but less than that seen with digitoxigenin.

Both **3b** and **4** did not markedly inhibit the enzyme; the latter result is consistent with the observation of Henderson and Chen¹⁷ that both 14,15- α - and 14,15- β epoxy derivatives of digitoxigenin are inactive in cardiotoxicity studies. The weak reversible activity seen in **2e** corresponds well with the report of Shigei, *et al.*,¹⁸ that 15 α -hydroxydigitoxigenin does not appreciably alter heart contractility. They have suggested that the 15 α -OH group may interfere with the stereochemical arrangement in the vicinity of the C and D rings.

The bromoacetates **2a**, **2b**, and **2c** all irreversibly inhibited the transport ATPase preparation to approxi-

(15) O. H. Lowry, N. J. Rosebrough, A. L. Farr, and R. J. Randall, J. Biol. Chem., 193, 265 (1951).

(16) K. Repke and H. J. Portius. Folia Haematol. (Leipzig). 83, 108 (1965).

(17) F. G. Henderson and K. K. Chen. J. Med. Chem. 8, 577 (1965).
(18) T. Shigei, M. Katori, H. Murase, and S. Imai. Experientia, 20, 572 (1964).

mately the same extent. Compound **3a** was also found to be a potent irreversible inhibitor of transport ATPase in spite of the fact that **3b** (which has very weak cardiotonic activity¹⁹) did not appreciably inhibit the enzyme. This is in contrast to the finding of Hokin, et al.,^{11a} that $\Delta^{3.14}$ -dianhydrostrophanthidin 3iodoacetate does not irreversibly inhibit transport ATPase whereas strophanthidin 3-iodoacetate does.

Experimental Section²⁰

3β-Hydroxy-14,15α-epoxy-5β,14α-card-20(22)-enolide 3-Acetate (4).—To 418 mg (1.1 mmoles) of **3b**¹³ in 8.6 ml of CHCl₃ was added a soln of 535 mg of *m*-ClC₆H₄CO₃H (purity 68%) in 13 ml of C₆H₆. The reaction mixt was allowed to stand for 27 hr at room temp. CHCl₃ (42 ml) was added followed by 16 ml of 10% Na₂SO₃ soln. The org layer was sepd and washed with 5% Na₂CO₃ soln and H₂O and after drying (Na₂SO₄) yielded 385 mg of residue. Recrystn of the residue from Me₃CO-Et₂O gave 376 mg (84% yield) of 4 as needles: mp 186-188° (lit.¹⁴ mp 220-226° from Me₂CO-petr ether; lit.²¹ mp 187-198° from MeOH-Et₂O); nmr δ 0.83 (3 H, s, 18-CH₃; calcd²² δ 0.82), 1.02 (3 H, s, 19-CH₃; calcd²² δ 1.01), 2.11 (3 H, s, 3-CH₃CO₄), 3.60 (1 H, m, 15-CH), 4.77 (2 H, q, J = 1 cps, 21-CH₄), 5.17 (1 H, m, 3-CH), 5.90 (1 H, q, J = 1 cps, 22-CH); mass spectrum, parent ion at *m/e* 414, a (P - H₃O) peak at *m/c* 396, and a (P - HOAc) peak at *m/e* 354.

 15_{α} -Hydroxydigitoxigenin 3-Acetate (2e).—This compd was prepd from 4 by the procedure described by Okada and Hasunuma.¹⁴ The product was obtained in 65% yield and after re-

(21) H. Ishii, T. Tozyo, and D. Satoh, Chem. Pharm. Bull., 11, 576 (1963).

(22) N. S. Bhacca and D. H. Williams, "Applications of NMR Spectroscopy in Organic Chemistry: Illustrations from the Steroid Field," Holden-Day, San Francisco, Calif., 1966, pp 14-24. crystn from MeOH-Et₂O had mp 260-262° (lit.¹⁴ mp 245-253° from Me₂CO-Et₂O; lit.²¹ mp 247-250° from MeOH-Et₂O).

15α-Hydroxydigitoxigenin 3-Acetate 15-Bromoacetate (2b).—A soln of 90 mg (0.2 mmole) of 2e in 3.6 ml of dry dioxane²³ and 2 drops of pyridine was treated with 2 drops of BrCH₂COBr, whereupon a granular ppt formed. The suspension was stirred at 70° for 16 hr. The reaction mixt was dild with 30 ml of H₂O, and 5% Na₂CO₃ soln was added to pH 7. The mixt was then concd to dryness, and the residue was purified by preparative tlc on silica gel (plates developed in CHCl₃-MeOH 10:1, major product had R_t 0.4) to give, after 1 recrystn from MeOH, 54 mg (47% yield) of 2b. An anal. sample had mp 230-232°; nmr δ 0.90 (3 H, s, 18-CH₃), 0.93 (3 H, s, 19-CH₃), 2.01 (3 H, s, 3-CH₃CO₂), and 3.80 (2 H, s, 15-BrCH₂CO₂). Anal. (C₄₇H₃₇-BrO₇) C, H, Br.

Digitoxigenin 3-Bromoacetate (2a).—A soln of 50 mg of digitoxigenin (5) in 2.5 ml of dry dioxane was treated with 2 drops of pyridine followed by 2 drops of BrCH₂COBr. The white suspension was stirred at room temp for 1.5 hr. The reaction mixt was dild with 10 ml of H₂O, and the ppt was filtered off, washed with H₄O, and dried *in vacuo*. Prep tlc of this solid on silica gel (plates developed in CHCl₃-MeOH, 96:4) gave 2 major bands. The lower band (R_1 0.2) yielded 10 mg of startingmaterial. The higher band (R_1 0.5) gave, after recrystn from MeOH, 38 mg (57% yield) of **2a**, mp 211–212°. Anal. (C₂₃H₃₅BrO₃) C, H.

 Δ^{14} -Anhydrodigitoxigenin 3-Bromoacetate (3a).—The reaction leading to this compd was carried out as described in the synthesis of 2a except that the reaction time was 3 hr. Prep tle of the crude product on silica gel (plates developed in CHCl₃-MeOH, 96:4) gave a major band at $R_{\rm f}$ 0.8 which, after recrystn from MeOH, gave 41 mg (65% yield) of 3a, mp 193–195°; mm δ 0.82 (3 H, s, 18-CH₃), 1.01 (3 H, s, 19-CH₃), 3.85 (2 H, s, 3-BrCH₂CO₂), and 5.27 (1 H, m, 15-CH). Anal. (C₂₅H₃₃BrO₄), C, H, Br.

Digoxigenin 3,12-Dibromoacetate (2c).—A solu of 78 mg of digoxigenin (6) in 3.6 ml of dry dioxane was treated with 2 drops of pyridine and 2 drops of BrCH₂COBr. The white suspension was stirred for 4 hr at 75°, dild with H₂O, and adjusted to pH 7 with 5% Na₂CO₃ soln. The solvents were removed under reduced pressure, and the residue was purified by prep tlc on silica gel (plates developed in CHCl₃-MeOH, 10:1). The major band at $R_1 0.7$ gave, after recrystn from MeOH-H₂O, 72 mg (56% yield) of **2c**, mp 215–218°. Anal. (C₂₇H₃₆O₄Br₂) C, H, Br.

Acknowledgments.—The authors are indebted to Dr. John Oliver's group for the nmr spectra and to Drs. Don DeJongh and David Brent for the mass spectral data. This work was made possible by a grant from the Michigan Heart Association.

(23) K. Hess and H. Frahm. Ber., 71, 2627 (1938).

β-Adrenergic Blocking Agents. 10. (3-Amino-2-hydroxypropoxy)anilides

A. F. CROWTHER, R. HOWE, AND L. H. SMITH*

Imperial Chemical Industries Limited, Pharmaceuticals Division, Alderley Park, Macclesfield, Cheshire, England

Received December 22, 1970

Several (3-amino-2-hydroxypropoxy) acylanilides have been synthesized. In experimental animals, they have potent β -adrenergic blocking actions on the myocardium but not at some other sites, for example, the peripheral blood vessels. Of the compounds tested 4-(2-hydroxy-3-isopropylaminopropoxy) acetanilide (practolol) was selected for clinical trial on the basis of optimal potency and selectivity.

In extension of our work on 1-amino-3-aryloxy-2propanols related to propranolol¹ we have prepared several analogs in which the aryl residue contains an acylamino substituent. In these preliminary studies, only relatively minor variations of the substituent (R_1) on the propanolamine side chain have been made. The acylamino substituents (R₂CONH) examined have included examples of alkanoyl, aroyl, and aralkanoyl groups. In general the compounds are potent β -adrenergic blocking agents. They differ, however, from previously known active compounds in that the inhibition of β -adrenergic responses is restricted to certain sites. Thus, the most studied compound of the new

⁽¹⁹⁾ K. Tokita, C. Isono, and Y. Kibayashi. Nippon Yakurigaku Zasshi. 58, 350 (1962).

⁽²⁰⁾ Melting points are corrected and were detd on a Fisher-Johns melting point apparatus. In spectra were detd with a Beckman Model IR-8 spectrophotometer and were consistent with the assigned structures. Nmr spectra were recorded with a Varian A-60A spectrometer in CDCla(MeiSi). In nmr descriptions s = singlet, q = quartet, m = multiplet. Mass spectra were taken on an A.E.I. MS-902 instrument using 70 eV with a direct source inlet system. Preparative tlc plates (0.75-mm thick), prepared using E. Merck silica gel G, were activated at 110° for 2 hr before use. Bands were located under uv light and were extd from the plates with MeOH-CHCls. 1:1. Elemental analyses were performed by Spang Microanalytical Laboratory, Ann Arbor, Mich. The anal. samples gave combustion analyses within 0.3% of theory.